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[Continued on nextpage]

(54) Title: NOVEL BACTERIAL STRAINS FOR BIOLOGICAL CONTROL OF MOSQUITOES

(57) Abstract: The present invention relates to novel bacteria strains that can be used in biological control against mosquito larvae (*Culex* spp.). The protein obtained from a novel *B.sphaericus* spp. isolates with the invention is used as larvicide, the step of isolating the protein at product obtaining stage is eliminated. By means of the invention, thee bacterial strains (MIB 5,6,7) in vestigated for biological control of mosquitoes are effective in both polluted and fresh water.



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# NOVEL BACTERIAL STRAINS FOR BIOLOGICAL CONTROL OF MOSQUITOES

### Field of the Invention

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The present invention relates to novel bacteria strains that can be used in biological control against mosquito larvae (*Culex* spp.).

#### **Background of the Invention**

- 10 Mosquitoes are vectors of many diseases such as Mosquito-borne arboviruses, malaria, filariasis and Japan encephalitis. Generally mosquito control does with chemical pesticides more than biopesticides in the world. These chemical pesticides are known as dichlorodiphenyltrichloro ethane (DDT), gammaxane, malathion, chlordane and organophosphates . All of them have high toxic range
- 15 for human health and environment. Compared to chemical pesticide, microbial insecticides are often species specific and do not contaminate environment, therefore, safe to non-target organisms in the nature. Among various microbial pesticides, *Bacillus thrungiensis* and *Bacillus sphaericus* are being widely used. Mosquitocidal bacteria are environmentally friendly alternatives to chemical 20 pesticides for controlling water mosquitoes.
- *Bacillus thrungiensis* subs. *israilensis* (Bti) is the most extensively used mosquito larvicidal bacteria in the world. Bti produces crystal glycoprotein (protoxin) coded by different genes such as Cry4A, Cry4B, Cry1OA, Cry11A and Cry1A during sporulation. Bti Cry toxins have been widely used in the control of broad range of
- 25 mosquito and blackfly species as well as nematodes, mite and protozoa. Another potential microbial pesticide insecticide, *Bacillus sphaericus*, is known to be effective against *Culex* spp. and *Anopheles* spp. species, and has better residual activity in polluted waters by production of binary toxin (Bin) and mosquitocidal toxins (Mtx). Mosquito resistance to some of *B. sphaericus* strains carrying a
- 30 single Bin (binary) toxin gene have been reported in many countries

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European Patent document no EP0349769, an application known in the state of the art, discloses *Bacillus sphaericus* bacteria genetically engineered with toxin producing genes taken from *Bacillus thuringiensis* var. *israelensis* (B.t.i.) bacteria and transferred to *Bacillus sphaericus* strains. The genetically modified (GM)

5 *Bacillus sphaericus* strains produced are capable of producing B.t.i. toxins in effective amounts and can control against mosquito larvae and black flies effectively.

European Patent document no EP0454485, an application known in the state of the art, discloses using insect killing toxins obtained from *Bacillus thuringiensis* or *Bacillus sphaericus* bacteria against pests living in water such as mosquito larvae. The spores of these bacteria kill some insect larvae feeding on these spores. The spores are digested in intestines of the larvae and release their toxins and neutralize the larvae. The known applications in the technique disclose taking

toxins of the bacteria to apply on larvae for biological control against mosquito.
Taking the toxins of the bacteria requires extra labor and cost. That is, protein isolation step is performed in these applications.

Even though, many commercial products are introduced to the market, development resistance in mosquito populations to some known biological control

20 products are always great need and force for the scientists to search for new natural mosquitocidal bacterial strains which can be used for development of new strains for development of new commercial microbial insecticide.

### Summary of the Invention

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The objective of the present invention is to provide novel bacterial strains that can be used as larvicide in biological control.

A further objective of the present invention is to provide novel bacterial strains for
biological control wherein the toxin protein isolation step in product obtaining
state is eliminated.

Another objective of the present invention is to provide novel bacterial strains for biological control which are effective in both polluted and fresh water.

### 5 **Detailed Description of the Invention**

"Novel Bacterial Strains for Biological Control of Mosquitoes" developed to fulfill the objective of the present invention is illustrated in the accompanying figures wherein,

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Figure 1 Bin 51 and bin 42 toxin genes PCR amplified by primers B.sph: B.sphaericus, MBI5, MBI6 and MBI7Figure 2 Mtx 1 and Mtx 2 toxin genes PCR amplified by primersB.sph: B.sphaericus, MBI5, MBI6 and MBI7

15 Figure 3 is the PCR bands of MBI5, MBI6 and MBI7 bacterial strains in gel imaging system (BIORAD) after electrophoresis in 1% agarose gel with ethidium bromide (NC: Negative Control).

Figure 4 Neighbour-joining tree: the phylogenetic relationships among the *Bacillus sphaericus-Yike* strains.

20 Figure 5 is the scanning electron microscope image of *B*. *sphaericus* bacterial cells.

Figure 6 is the scanning electron microscope image of MBI5 bacterial strains. Figure 7 is the scanning electron microscope image of MBI6 bacterial strains. Figure 8 is the scanning electron microscope image of MBI7 bacterial strains.

Figure 9 is the 16 rDNA sequence of MBI5 bacterial strain.Figure 10 is the 16S rDNA sequence of MBI6 bacterial strain.Figure 11 is the 16S rDNA sequence of MBI7 bacterial strain.

In the inventive biological control against the mosquito larvae, the strains of *B*. 30 *sphaericus* species are applied against the mosquito larvae. Deposit number is taken for the inventive strains from United States Department of Agriculture

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Research, Education and Economics Agricultural Research Service on January 28, 2009. The deposit numbers of sub strains belonging to *B. sphaericus* species and named MBI5, MBI6, MBI6 are respectively registered as NRRL B-50199, NRRL B-50200 and NRRL B-50201.

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In laboratory experiments carried out against mosquito larvae (*Culex* spp.), it has been found out that the larvicide effects of *B.sphaericus* MIB5,6,7 strains and presence of Bin genes are the same as in the commercial strains of *B. sphaericus*. Furthermore, it has been observed that in experiments against the mosquito larvae

- 10 the inventive bacteria strains show faster effect in a higher ratio than the known *B.sphaericus* strains. In the applications of the previous technique, an extra process is performed in order to obtain protein from the isolates. By means of the invention, the protein isolation step in obtaining product stage is eliminated. It is observed that newly found bacteria strains (MBI5, MBI6, MBI7) are effective
- 15 when they are given to the medium in which the larvae are present directly without performing protein isolation. At the same time *Bacillus sphaericus* strains show high larvicide effect both in polluted and fresh water. Various experimental studies have been carried out in order to test the effectiveness of the invention.

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### **Experimental Studies**

25 Single colonies of newly isolated bacterial strains and Bti 4Q4, Bti ATCC 35646, *B. sphaericus* and were cultivated on to NYSM (Nutrient Yeast Salt Medium) agar and incubated for 48h at 30°C. Bacterial growth of each strain was harvested and resuspended in 10 ml of distilled water. Absorbance was adjusted to 0.2 with water and then 1 ml of suspension was added to 100 ml of fresh water/polluted

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water in 250ml flasks containing 100 larvae (at the stage of 3 or 4<sup>th</sup> instar) of *Culex* spp. The inoculated flasks were maintained on laboratory bench and observed for 48h at room temperature. In order to determine larvicidal bacterial strains, which were capable of killing 90% of larvae, positive and negative control flasks treated with reference strains and sterile water, respectively, were kept the same condition as inoculating ones. After toxicity test, three strains of *B.sphaericus* (MBI 5, 6, 7) were selected as high toxic mosquitocidal bacteria and used for further studies. According to the bioassay test results MBI 5,6,7 have

three bacteria were done in fresh and polluted water that contained 100 larvae (see
 Table 1). Bti ATCC 35646, Bti 4Q4 and commercial *B.sphaericus* were used as .
 positive control.

potential to be toxic to larvae of *Culex* spp. Investigation of larvacidal features of

Table 1: The effectiveness of MBI 5, MBI 6, MBI 7 strains and *B.sphaericus*, Bti
ATCC 35646 and Bti 4Q4 bacteria against *Culex* spp. larvae in polluted and fresh water.

Bacteria name		<i>Culex</i> spp. Live larvae number (500ml water/100 live larvae)		
		Pollute	Fresh water	
		24h	48h	24h
B.sphaericus		10	4	0
(500µL)	i			
MBI	5	6	4	0

(500µL)			
MBI 6	7	3	0
(500µL)			
MBI7	9	2	0
(500µL)			
BtiATCC35646	20	20	16
(500µL)			
Bti 4Q4 (500μL)	34	32	24

According to the test results, it was found that MBI 5, MBI 6, MBI 7 strains are more effective in polluted water in 24 hours relative to the known *B.sphaericus*, Bti ATCC 35646 and Bti 4Q4 bacteria. The effectiveness percentage of MBI 5, MBI 6, MBI 7 strains were determined as 94%, %93 and %91, respectively. In tests performed in fresh water, it was observed that MBI 5, MBI 6, MBI 7 strains and *B.sphaericus* bacteria have 100% success by killing all existing healthy larvae in 24 hours. On the other hand, it was found out that Bti ATCC 35646 and Bti 4Q4 bacteria are effective against larvae in ratio of 84% and 76% in fresh water, respectively (Table 1).

#### **Diagnostic studies**

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#### Phenotypic diagnostic studies

15 All of the methods provided to understand cell properties of three new strains of bacillus. MBI5, MBI6 and MBI7 were an aerobic, Gram-positive bacteria According to electron microscope images of MBI5, MBI6, MBI7 and *B.sphaericus*, they are rod-shaped bacteria (Figure-6, Figure-7, and Figure-8) and similar with *B.sphaericus* (Figure-5).

They were also growth 20-35°C, and the optimum growth temperatures were 27-30°C. Growth at 50°C and 4°C were not observed on nutrient agar. The physiological characteristics of MBI5, MBI6 and MBI7 were summarized and selective characteristics with related model as *B.sphaericus* were compared (Table

5 2).

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**Table 2:** Phenotypic characteristics of strains MBI5, MBI6, MBI7 compared

 with commercial *B.sphaericus*.

	Characteristics	MBI5	MBI6	MBI7	B.sphaericus
15	Gram staining	+	+	+	+
	Oxidase	-	-	-	-
	Catalase	-	-	· -	-
	Capsule Staining	+	+	+	+
	Endospor Staining	+	+	+	+
20	Hemolysis	+	+	+	+
	Anaerobic test	-	-	-	-
	Penicilline	+	+	+	+

(+, positive; -, negative)

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### Fatty acid profile analysis

Each MBI strains were characterized as unique and novel in terms of BIOLOG, FAME profiles and 16S rRNA sequencing data.

30 The cellular fatty acid profiles of MBI 5, 6, 7 and *B.sphaericus* were listed in Table 3. The major cellular fatty acids in MBI5 included iso-pentadecanoic acid (C i5:0 iso, 45,00%) and Ci<sub>6</sub> 0 iso, 12,65%. Minor amounts of the iso-branched fatty

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acids  $Ci_{4;0}$  iso (0.60%),  $C]_{6:0}(1.72\%)$ ,  $C_{7:}$ , iso  $\omega 1 \propto (1,43\%)$ . The major cellular fatty acids in MBI6 included iso-pentadecanoic acid ( $Ci_{5:0}$  iso, 44,99%) and  $C|_{6:0}$  iso, 15,24%. Minor amounts of the fatty acids  $Ci_{6:0}$  (0.78%),  $C_{17:1}$  iso colOc (1,40%). The major cellular fatty acids in MBI7 included iso-pentadecanoic acid

- 5 (C<sub>15:0</sub> iso, 45,84%) and C<sub>15:0</sub> anteiso, 13,13%. Minor amounts of the iso-branched fatty acids C|4.0 iso (0.68%), Ci<sub>8:</sub>i iso co9c (1,03%). Consequently, significant similarities in fatty acids profiles were found between *B.sphaericus* and MBI group. All of the groups MBI and *B.sphaericus* were identified with MIDI as *Bacillus-sphaericus-* GC subgroup E.
- 10 Table 3: Cellular fatty acid composition of MB1 5, 6, 7 and B.sphaericus

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Numerical Names	Percentage	Percentage	Percentage	Percentage %
of the Fatty acids	%	%	%	B.sphaericiis
(Peak names)	MBI 5	MBI 6	MBI 7	
14:0 iso	2,02	4,38	1,51	1,26
14:0	0,60	-	0,68	0,85
15:0 iso	45,00	44,99	45,84	46,61
15:0 anteiso	10,87	9,22	13,13	7,89
14:0 iso 30H	-	-	-	1,05
16:1 w7c alkol	9,93	12,38	9,55	6,80
16:iso	12,65	15,24	8,14	5,48
16:1 wile	3,31	2,04	3,31	5.62
16:0	1,72	0,78	1,78	1,64
17:1 iso wlOc	1,43	1,40	2,35	4,92
Sum In Feature 4	1,65	1,72	2,32	2,58
17:0 iso	6,1 1	4,67	5,69	10,86
17:0 anteiso	4,70	3,19	4,67	4,45
18:1 w9c	-	-	1,03	-
Summed Feature 4	1,65	1,72	2,32	2,58

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Sequence analysis with nucleic acid based 16S-rDNA PCR amplification DNA extraction from bacterial strains:

Total genomic DNA from bacterial strains was extracted according to methodology described by Jimenez with some modifications. The pure strains were

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cultured in Nutrient Agar (NA) solid medium 16-20 hours at 27C and one single colony contaminated into 10ml Nutrient Broth (NB) at 27C for 3-4 hours until the absorbances up to 1 at 660nm. The bacterial cells were collected from media afer 10min at 2000g centrifugation. The cells were suspended with lml of Tris-EDTA

5 buffer(10mM Tris Base, IraM EDTA, 0.05% Tween 20, pH 9.0) and transfered into 2ml microcentrifuge tube, centrifuged at 14000g for 2min, supernatant discarded from the tube and added lml of Tris-EDTA buffer and repeated application for 3 times. Finally 300µ1 of Tris-EDTA buffer added and boiled at 94C for 30 min in water bath. Centrifuged at 14000g for 2min and 200µ1 DNA

10 was collected from supernatant and stored at -20 for further PCR applications.
PCR amplification and purification of 16S rRNA:
16S rRNA genes of the bacterial DNA isolates (MBI 5, MBI 6, MBI 7 and *Bacillus sphaericus* serotype H for control) amplified by the PCR (BIORAD, İtaly) using purified DNA and primers 27f and 1492r (Lane, 1991). PCR

- 15 amplifications was caried out in total volume of 50ul reaction mixture containing 0.2 mM of 27f and 1492r primers for total 16S, 1 U of *pfu* DNA polymerase (Fermentas, USA), 0.2mM of each deoxynucleoside triphosphate (dNTP), 1 mM MgS04, 10mM Tris and 50ng template DNA. PCR conditions were as follows : preamplification 94°C for 5 min : denaturation at 94°C for 30s : annealing at 55°C
- for 40s : elongation at 72°C for 2min repeated 34 cycles and then post amplification for final extention 10 min at 72°C.
  We designed specific two new primers for *Bacillus sphaericus* like members of Bacillaceae family. We amplified 550bp of 16S rRNA gene fragments of the bacterial DNA isolates (MBI 5, MBI 6, MBI 7 and *Bacillus sphaericus* serotype H
- for control) by the PCR (BIORAD, Italy) using purified DNA and primers FAMI and FAM2. PCR amplifications was caried out in total volume of 50ul reaction mixture containing 0.2 mM of FAM1 and FAM2 primers for 550bp of 16S, 1 U of *pfu* DNA polymerase (Fermentas, USA), 0.2mM of each deoxynucleoside triphosphate (dNTP), 1 mM MgS04, 10mM Tris and 50ng template DNA. PCR
  conditions were as follows : preamplification 94°C for 5 min : denaturation at

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94°C for 30s : annealing at 51°C for 40s : elongation at 72°C for 45sec repeated 34 cycles and then post amplification for final extention 10 min at 72°C.

The amplified DNA products was detected by using Biorad image analysing system (BIORAD, İtaly) after electrophoresis of PCR amplicons in a 1% agarose

5 gel stained with ethidium bromide.

16S rRNA gene sequencing and phylogenetic analysis

Pure amplification products were sequenced with a Prism ABI 3100 Genetic Analyzer 16 caillaries, dideoxy terminator cycle sequencing kit (Applied Biosystems). The protocols used were due to manufacturers recommendations.

- 10 Sequences were determined with an automated DNA sequencer (model: Prism ABI 3100; Applied Biosystems). Both strands were sequenced using the primers 27f, 1492r, FAM1 and FAM2 (Lane, 1991; Nakamura, 1996). The clustal w program (Higgins *et al.*, 1992) was used to align the 16S DNA sequences generated with sequences of *Bacillus sphaericus* like members from GenBank
- 15 NCBI (Larsen *et al.*, 1993). The sequences of 16s rDNA genes were obtained (Figure-9, Figure-10, Figure-11).

Genetic distance was computed by using Kimura's two-parameter model (Kimura, 1980) and used for neighbour-joining analysis. Phylogenetic trees were constructed using neighbour-joining and maximum-parsimony methods provided

20 by CLC Genomics Workbench\_2\_1\_1 both methods produced trees with similar topologies. Nucleotide sequences generated in this study have been deposited with GenBank under the accession numbers.

Another study was Neighbour-joining tree analysis that is based on 1450 nucleotide sequences. Confidence limits estimated from bootstrap analyses (100

- 25 replications) appear at the nodes. A maximum-parsimony tree generated from the sequence data exhibited similar topology to this tree. In the phylogenetic tree; MBI5, MBI6 and MBI7 clearly belonged to the strains of *Bacillus sphaericus*, as shown by the high bootstrap value (Figure-4).
- 30 Determination of Toxin Genes

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Toxin genes investigated according to methodology described by Nishiwaki et al., PCR of toxin genes of the bacterial DNA isolates (*MBI 5, MBI 6, MBI 7* and *Bacillus sphaericus serotype H* for control) possesses has done for the genes encoding the mosquitocidal binary toxin (51 and 42 kDa), Mtx1, and Mtx2. PCR was constructed according to the following conditions: preamplification 94°C for 2 min followed by 30 cycles of denaturation at 94°C for 15 s, annealing at 55°C for 30 s, and elongation at 72°C for 1 min 30 s. The master mix consisted of 1 U

of TSG polymerase (Biobasic,Canada), 1 mM MgS04, 0.2 mM of each deoxynucleoside triphosphate (dNTP), 20 ng of template DNA, and 5 pmol of each primer in total volume of 50ul reaction mixture.

The amplified DNA products was detected by using Biorad image analysing system (BIORAD, İtaly) after electrophoresis of PCR amplicons in a 1% agarose gel stained with ethidium bromide (Figure-1, Figure-2).

The PCR amplification of Bin and Mtx toxin genes of MBI 5, 6, 7 and

commercial *B. Sphaericus* have done. Figure 1 reveals that *B. sphaericus*, MBI 5, MBI 6 and MBI 7 have Bin 51 and Bin 42 toxins. At the same time, MBI 5, MBI 6 and MBI 7 have not Mtx land Mtx 2 toxins (Figure-2). In addition, commercial *B. sphaericus* has both Bin and Mtx toxins.

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#### CLAIMS

**1.** MBI 5, MBI 6, MBI 7 bacterial strains which are sub strains of *Bacillus sphaericus* bacteria and which are used in biological control.

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**2. A** MBI 5 bacteria strain for biological control according to claim 1, which is deposited with NRRL B-50199 number.

3. A MBI 6 bacteria strain for biological control according to claim 1, which isdeposited with NRRL B-50200 number.

4. A MBI 7 bacteria strain for biological control according to claim 1, which is deposited with NRRL B-50201 number.

15 **5.** Bacterial strains for biological control according to claim 1 to 4, which are effective against mosquito larvae.

**6.** Bacteria strains for biological fight according to claim 5, which are effective when they are given directly to the medium in which the larvae are present without isolating protein.

7. Bacterial strains for biological control according to claim 6, which shows high larvicide effect in fresh and polluted water.

25 8. Bacteria strains for biological contol according to claim 7, which are an aerobic, Gram-positive, rod-shaped bacteria

9. Bacterial strains for biological control according to claim 8, which have 99% closeness with *Bacillus* sp. ZYM and *Bacillus* sp. BD-95.

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**10.** Bacterial strains for biological control according to claim 9, which includes Bin 51 and Bin 42 toxin genes.









1500 bp











<u>MBI 5</u>
CGAATAATCIGTTICACCICAIGGIGAAACACIGAAAGACGGITICGGCIGICGCIATA
GGATGGGCCCGCGCGCGCATTAGCTAGTTGGTGAGGTAACGGCTCACCAAGGCGACGAT
GCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGCCCAGACTC
GGCGAAAGCCTGATGGAGCAACGCCGCGTGAGTGAAGAAGGATTTCGGTTCGTAAAAC
TCIGIIGIAAGGGAAGAACAAGTACAGTAGTAACIGGCIGIACCTIGACGGTACCTIAT
TAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGT
TGTCCGGAATTATTGGGCGTAAAGCGCGCGCAGGT
GGTTICTTAAGICTGATGIGAAAGCCCACGGCTCAACCGTGGAGGGTCATTGGAAACTG
GGAGACTTGAGTGCAGAAGAGGATAGTGGAATTCCAAGTGTAGCGGTGAAATGCGTAG
AGATTTGGAGGAACACCAGTGGCGAAGGCGACTATCTGGTCTGTAACTGACACTGAGGC
GCGAAAGCGTGGGGGGGGCAAACAGGATTAGATACC
CTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGGTTTCCGCCCCTTAGT
GCTGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCA
AAGGAATTGACGGGGGCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACG
CGAAGAACCTTACCAGGTCTTGACATCCCGTTGA
CCACTGTAGAGATATAGTTTCCCCTTCGGGGGGCAACGGTGACAGGTGGTGCATGGTTGT
CGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTGATCT
TAGTTGCCATCATTTAGTTGGGCACTCTAAGGTGACTGCCGGTGACAAACCGGAGGAAG
GTGGGGATGACGTCAAATCATCATGCCCCTTAT
GACCTGGGCTACACGTGCTACAATGGACGATACAAACGGTTGCCAACTCGCGAGAG
GGAGCTAATCCGATAAAGTCGTTCTCAGTTCGGATTGTAGGCTGCAACTCGCCTACATG
AAGCCGGAATCGCTAGTAATCGCGGATCAGCATGCCGCGGTGAATACGTTCCCGGGCCT
TGTACACCGCCCGTCACACCACGAGAGTTTGT
AACACCCGAAGTCGGTGAGGTAACCTTTGGAGCCAGCCGCCGAAGGTGGATAGATGAT

# <u>MBI 6</u>

TGCAAGTCGAGCGAACAGAGAAGGAGCTTGCTCCTTCGACGTTAGCGGCGGACGGG TGAGTAACACGTGGGCAACCTACCTTATAGTTTGGGATAACTCCGGGAAACCGGGG CTAATACCGAATAATCTGTTTCACCTCATGGTGAAACACTGAAAGACGGTTTCGGCT GTCGCTATAGGATGGGCCCGCGCGCGCATTAGCTAGTTGGTG AGGTAACGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCA CACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTC TCGTAAAACTCTGTTGTAAGGGAAGAACAAGTACAGTAGTAA CTGGCTGTACCTTGACGGTACCTTATTAGAAAGCCACGGCTAACTACGTGCCAGCAG CCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGC GCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCCACGGCTCAACCGTGGAGGGTCATT GGAAACTGGGAGACTTGAGTGCAGAAGAGGATAGTGGAAT TCCAAGTGTAGCGGTGAAATGCGTAGAGATTTGGAGGAACACCAGTGGCGAAGGCG AGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGGTTTCCG CCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTGG GGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCGG TGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACCTTACCAGGTCTTGACATCC CGTTGACCACTGTAGAGATATAGTTTCCCCTTCGGGGGGCAACGGTGACAGGTGGTGC ATGGTTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAG TCCCGCAACGAGCGCAACCCTTGATCTTAGTTGCCATCATTTAGTTGGGCACTCTAA GGTGACTGCCGGTGACAAACCGGAGGAAGGTGGGGGATGACGTCAAATCATCATGCC CCTTATGACCTGGGCTACACACGTGCTACAATGGACGATACAAACGGTTGCCAACTC GCGAGAGGGGGGGGCTAATCCGATAAAGTCGTTCTCAGTTCGG ATTGTAGCTGCAACTCGCCTACATGAAGCCGGAATCGCTAGTAATCGCGATCAGCAT GCCGCGGTGAATACGTTCCCGGGCCTTGTA

## <u>MBI 7</u>

AGGAGCTTGCTCCTTCGACGTTAGCGGCGGACGGGTGAGTAACACGTGGGCAACCTA CCTTATAGTTTGG GATAACTCCGGGAAACCGGGGCTAATACCGAATAATCTGTTTCACCTCATGGTGAAAC GTGAGGTAACGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGC CACACTGGGACTGAGACACGGCCCAGACTCCTACGGG AGGCAGCAGTAGGGAATCTTCCACAATGGGCGAAAGCCTGATGGAGCAACGCCGCGT GAGTGAAGAAGGATTTCGGTTCGTAAAACTCTGTTGTAAGGGAAGAACAAGTACAGT AGTAACTGGCTGTACCTTGACGGTACCTTATTAGAAAGCCACGGCTAACTACGTGCCA GCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGG AATTATTGGGCGTAAAGCGCGCGCGCGGGTGGTTTCTTAAGTCTGATGTGAAAGCCCACG GCTCAACCGTGGAGGGTCATTGGAAACTGGGAGACTTGAGTGCAGAAGAGGATAGTG GAATTCCAAGTGTAGCGGTGAAATGCGTAGAGATTTGGAGGAACACCAGTGGCGAAG GCGACTATCTGGTCTGTAACTGACACTGAGGCGCGAAA GCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGTG CTAAGTGTTAGGGGGGTTTCCGCCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGC CTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGCCCGCACAAG CGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGA ACCTACCAGGTCTGACTTCCCGTT

International application No PCT/IB2012/053426

A. CLASSIFICATION OF SUBJECT MATTER INV. A01N63/00 A01P7/04 ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

A01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, BIOSIS, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT				
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X Furth	er documents are listed in the continuation of Box C.	X See patent family annex.		
* Special ca "A" document to be o "E" earlier a filing da "L" document cited to special "O" document means "P" document the prio	ategories of cited documents : t defining the general state of the art which is not considered f particular relevance pplication or patent but published on or after the international ate twhich may throw doubts on priority claim(s) orwhich is o establish the publication date of another citation or other reason (as specified) tt referring to an oral disclosure, use, exhibition or other t published prior to the international filing date but later than rity date claimed	<ul> <li>"T" later document published after the intern date and not in conflict with the applicat the principle or theory underlying the in"X" document of particular relevance; the cl. considered novel or cannot be consider step when the document is taken alone</li> <li>"Y" document of particular relevance; the cl. considered to involve an inventive step combined with one or more other such being obvious to a person skilled in the</li> <li>"&amp;" document member of the same patent fit."</li> </ul>	ational filing date or priority ion but cited to understand evention aimed invention cannot be ed to involve an inventive aimed invention cannot be when the document is documents, such combination art	
Date of the a	actual completion of the international search	Date of mailing of the international sear	ch report	
20	0 November 2012	03/12/2012		
Name and m	nailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Authorized officer Habermann , Jbrg		

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×	MULLIGAN III F S ET AL: "Laboratory and Field Evaluation of Bacillus sphaericus as a Mosquito Control Agent", JOURNAL OF ECONOMIC ENTOMOLOGY, ENTOMOLOGICAL SOCIETY OF AMERICA, LANDHAM, MD, US, vol. 71, no. 5, 1 October 1978 (1978-10-01), pages 774-777, XP001471073, ISSN: 0022-0493 page 774, column 2, line 32 - line 42 page 775, column 1, line 20 - line 33 page 775, column 2, line 12 - line 15 page 776, column 1, line 19 - line 42	1-10
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Information on patent family members

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